

## A DITERPENE RELATED TO ERYTHROXYDIOL FROM *HELICHRYSUM REFLUXUM*

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**Key Word Index**—*Helichrysum refluxum*; Compositae; diterpene; erythroxane derivative.

**Abstract**—From *Helichrysum refluxum*, in addition to known compounds, a diterpene acid related to erythroxydiol Z was isolated. The structure was elucidated by NMR techniques.

In continuation of our investigation of *Helichrysum* species [1], we have now studied a species from Transvaal, *H. refluxum* N.E.Br. The aerial parts afforded in addition to widespread compounds several known diterpenes belonging to the kaurane, trachylobane and stachane series (see Experimental) as well as a new diterpene, the acid 1, which was isolated as its methyl ester. The carbon skeleton was that of some rare diterpenes which have been isolated from an *Erythroxylon* species [2]. In the  $^1\text{H}$  NMR spectrum of the ester in deuteriobenzene (Table 1) nearly all the signals could be assigned by spin decoupling thus clearly indicating that methyl groups were at C-5, C-9 and C-13. Irradiation of the low-field double-doublet at  $\delta 6.64$ , which obviously was that of H-3,

allowed the assignment of H-2 $\alpha$  and H-2 $\beta$ . As the latter were further coupled with the protons which showed signals at  $\delta 1.50$  and 1.36, the H-1 signals were assigned as well. As these signals were coupled with a double-doublet at  $\delta 1.03$ , a proton was at C-10 which, however, had no further vicinal protons. Furthermore, the sequence H-6–H-7–H-8–H-14 as well as the signals of H-11 and H-12 could be fully assigned by spin decoupling. The configuration at C-13 was supported by the observed W-coupling of H-12 $\alpha$  with H-17 which itself could be assigned by NOE difference spectroscopy. Irradiation of H-17 showed clear NOEs with H-15 and H-16 *trans*. Irradiation of H-19 and H-20 gave NOEs with H-7 $\alpha$ . Accordingly, both methyls were axially orientated. The

Table 1.  $^1\text{H}$  NMR spectral data of 1-methyl ester (400 MHz, TMS as internal standard)

	$\text{C}_6\text{H}_6$	$\text{CDCl}_3^*$		$\text{C}_6\text{D}_6$	$\text{CDCl}_3$
H-1 $\alpha$	1.38 <i>dd</i>		H-12 $\alpha$	1.63 <i>br ddd</i>	
H-1 $\beta$	1.50 <i>dd</i>	1.73 <i>dd</i>	H-12 $\beta$	1.22 <i>ddd</i>	
H-2 $\alpha$	2.01 <i>dddd</i>	2.30 <i>dddd</i>	H-14 $\alpha$	1.36 <i>dd</i>	
H-2 $\beta$	1.91 <i>dddd</i>	2.16 <i>dddd</i>	H-14 $\beta$	0.99 <i>dd</i>	
H-3	6.64 <i>dd</i>	6.56 <i>dd</i>	H-15	6.38 <i>dd</i>	6.40 <i>dd</i>
H-6 $\alpha$	2.7 <i>ddd</i>	2.34 <i>ddd</i>	H-16 $\alpha$	4.98 <i>dd</i>	4.84 <i>dd</i>
H-6 $\beta$	1.35 <i>ddd</i>		H-16 $\beta$	5.05 <i>dd</i>	4.91 <i>dd</i>
H-7 $\alpha$	1.44 <i>dddd</i>	1.46 <i>m</i>	H-17	1.05 <i>br s</i>	1.00 <i>s</i>
H-7 $\beta$	1.15 <i>dddd</i>		H-19	1.47 <i>s</i>	1.26 <i>s</i>
H-8	1.35 <i>dddd</i>		H-20	0.70 <i>s</i>	0.75 <i>s</i>
H-10	1.00 <i>dd</i>	1.13 <i>dd</i>	OMe	3.48 <i>s</i>	3.68 <i>s</i>
H-11 $\alpha$	1.44 <i>ddd</i>	1.44 <i>m</i>			
H-11 $\beta$	0.89 <i>ddd</i>				

\*Remaining signals are overlapped multiplets.

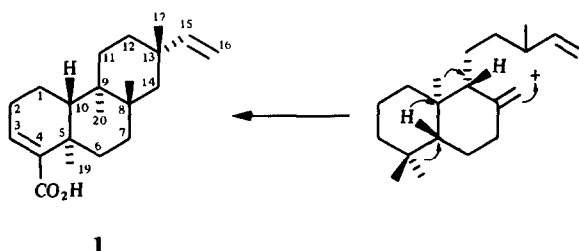
$J$  (Hz): 1 $\alpha$ , 1 $\beta$  = 13.5; 1 $\alpha$ , 2 $\alpha$  = 1.5; 1 $\alpha$ , 2 $\beta$  = 10; 1 $\alpha$ , 10 = 13; 1 $\beta$ , 2 $\alpha$  = 5; 1 $\beta$ , 2 $\beta$  = 7; 2 $\alpha$ , 2 $\beta$  = 19; 2 $\alpha$ , 3 = 4.5; 2 $\beta$ , 3 = 3; 6 $\alpha$ , 6 $\beta$  = 12.5; 6 $\alpha$ , 7 $\alpha$  = 6 $\alpha$ , 7 $\beta$  = 3; 6 $\beta$ , 7 $\alpha$  = 11; 6 $\beta$ , 7 $\beta$  ~ 3; 7 $\alpha$ , 7 $\beta$  = 13; 7 $\alpha$ , 8 = 10; 7 $\beta$ , 8 ~ 3; 8, 14 $\alpha$  = 11; 8, 14 $\beta$  = 3; 11 $\alpha$ , 11 $\beta$  = 13; 11 $\alpha$ , 12 $\alpha$  = 4; 11 $\alpha$ , 12 $\beta$  ~ 3; 11 $\beta$ , 12 $\alpha$  = 13; 11 $\beta$ , 12 $\beta$  = 4; 12 $\alpha$ , 12 $\beta$  = 13; 12 $\alpha$ , 17 ~ 0.5; 15, 16 $\alpha$  = 11; 15, 16 $\beta$  = 17.5; 16 $\alpha$ , 16 $\beta$  = 1.5.

$^{13}\text{C}$  NMR spectrum was in agreement with the proposed structure and configuration (see Experimental). The observed optical rotation may be an indication that **1** has the same absolute configuration as erythrodiol [2]. We propose for this carbon skeleton the name erythropane; **1** therefore is erythroxa-3,15-dien-18-oic acid. Most likely this system is formed as indicated in Scheme 1. *Helichrysum* species which contain diterpenes of different types have been observed previously [1], though normally other constituents were isolated.

#### EXPERIMENTAL

The air-dried plant material (voucher 81/123, collected in February 1981 in Transvaal) was worked-up in the usual fashion [3]. The crude extract of the aerial parts (175 g) was first separated by CC affording the following fractions: 1 (petrol), 2 ( $\text{Et}_2\text{O}$ -petrol, 1:9), 3 ( $\text{Et}_2\text{O}$ -petrol, 1:3) and 4 ( $\text{Et}_2\text{O}$ -petrol, 1:1 and  $\text{Et}_2\text{O}$ ). TLC of fraction 1 (petrol) gave 7 mg bicyclogermacrene, 4 mg cadinene, 4 mg aromadendrene, 3 mg  $\alpha$ -cedrene and 7 mg squalene (identified by GC/MS and by  $^1\text{H}$  NMR comparison with authentic material). TLC of fractions 2 and 3 ( $\text{Et}_2\text{O}$ -petrol, 1:3) gave 5 mg trachylobanic acid, 10 mg *ent*-kaurenic acid, 100 mg beyeren-19-oic-acid [4], 70 mg lupeol and its  $\Delta^{12}$ -isomer (*ca* 1:2) and 40 mg crude **1** which was purified as its methyl ester by TLC ( $\text{Et}_2\text{O}$ -petrol, 1:9) ( $R_f$  0.6). TLC of fraction 4 ( $\text{Et}_2\text{O}$ -petrol, 1:1) gave 75 mg oleanolic acid, 20 mg 2,3-dihydroaromaticin [5] and 15 mg carabron. [6]. Known compounds were identified by comparing the 400 MHz  $^1\text{H}$  NMR spectra with those of authentic material and by co-TLC in different solvent systems.

*Methyl erythroxa-3,15-dien-18-oate*. Colourless crystals, mp  $103^\circ$ ;  $\text{IR}_{\text{max}}^{\text{CCl}_4}$   $\text{cm}^{-1}$ : 1720 ( $\text{CO}_2\text{R}$ ), 915 ( $\text{CH}=\text{CH}_2$ ); MS  $m/z$  (rel. int.): 316.240  $[\text{M}]^+$  (91) (calc. for  $\text{C}_{21}\text{H}_{32}\text{O}_2$ : 316.230), 301  $[\text{M}-\text{Me}]^+$  (85), 285  $[\text{M}-\text{OMe}]^+$  (64), 269  $[301-\text{MeOH}]^+$  (11), 257  $[285-\text{CO}]^+$  (33), 242  $[269-\text{CH}=\text{CH}_2]^+$  (56), 107  $[\text{C}_8\text{H}_{11}]^+$  (100);  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , C-1-C-20): 16.9 *t*, 27.3 *t*, 136.7 *d*, 142.5 *s*, 38.0 *s*, 32.0 *t*, 25.8 *t*, 42.4 *d*, 36.4 *s*, 36.7 *t*, 35.1 *t*,



Scheme 1.

36.8 *s*, 39.0 *t*, 151.3 *d*, 108.6 *t*, 23.2 *q*, 167.9 *s*, 21.5 *q*, 12.4 *q*, 54.1 *q* (OMe);

$$[\alpha]_{24}^{25} = \frac{589}{-84} \frac{578}{-104} \frac{546}{-125} \frac{436 \text{ nm}}{-225} (\text{CHCl}_3; c 0.5).$$

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